

Aug 09, 2019

Thiobarbituric acid reactive substances (TBARS) Assay

DOI

dx.doi.org/10.17504/protocols.io.3sngnde



Eva Feldman¹

¹University of Michigan - Ann Arbor

Diabetic Complications Consortium
Tech. support email: rmcindoe@augusta.edu



Lili Liang

OPEN  ACCESS



DOI: dx.doi.org/10.17504/protocols.io.3sngnde

External link: <https://www.diacomp.org/shared/document.aspx?id=33&docType=Protocol>

Protocol Citation: Eva Feldman 2019. Thiobarbituric acid reactive substances (TBARS) Assay. [protocols.io](https://dx.doi.org/10.17504/protocols.io.3sngnde)
<https://dx.doi.org/10.17504/protocols.io.3sngnde>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: June 05, 2019

Last Modified: August 09, 2019

Protocol Integer ID: 24110

Keywords: Biochemical Measures of Neuropathy, diabetic neuropathy, TBARS

Abstract

Summary:

Plasma concentrations of thiobarbituric acid reactive substances (TBARS) are an index of lipid peroxidation and oxidative stress. The protocol describes how the DiaComp quantitates TBARS in the animal models.

Diabetic Complication:



Neuropathy

Materials

MATERIALS

⊗ Thiobarbituric Acid (TBA) **Catalog #ICN 190284**

⊗ Trichloroacetic Acid **Sigma Aldrich Catalog #490-10**

⊗ 1133-tetramethoxypropane **Acros Organics Catalog #148611000**

Reagent Preparation:

Thiobarbituric Acid (TBA): 67 mg in 1mL DMSO then add 9 mL H₂O.

10% Trichloroacetic Acid (w/v): in H₂O.

1,1,3,3-tetramethoxypropane: 4.167 μL in 1mL Ethanol then add 49 mL H₂O. (500 μM)

Note:

Sigma-Aldrich RRID:SCR_008988

Sample Preparation:

1 Plasma:

- Place 100µL plasma into a labeled 1.5mL micro-centrifuge tube.

Tissue:

- Label 1 sets of 1.5mL micro-centrifuge tubes, 1 set screw top tubes and 1 set of 0.5mL tubes.
 - Weighed out ~20mg and sonicate in 200µL RIPA buffer + inhibitors.
 - Sonicate.
 - Centrifuged @ 3000 for 10 min @ 4° .
1. Remove 10 µL aliquot into the 0.5mL tubes for protein analysis.
 2. Place 100 µL lysate into a labeled 1.5mL micro-centrifuge tube.
 3. Add 200µL ice cold 10% Trichloroacetic acid to precipitate protein.
 4. Incubate for 15 minutes on ice.
 5. Prepare standards as follows:

CONCENTRATION (µM)	H ₂ O	TETRAMETHOXYPROPANE
0	500	-----
0.625	500	500 from tube 3
1.25	500	500 from tube 4
2.5	500	500 from tube 5
5.	500	500 from tube 6
10	800	200 from tube 7
50	500	500 from tube 8
100	800	200 of 500uM stock

6. Centrifuge samples @ 2200 x g for 15 min. at @ 4°C.



7. Place 200 μ L supernatant and standards into new labeled screw top 1.5ml tube.
8. Add and equal volume of 0.67% (w/v) TBA.
9. Incubate in a boiling water bath for 10 min.
10. Cool. Sample is ready for assay.

Performing Assay:

- 2
 1. While samples are cooling, layout on computer and save as TBxxxxxx.sed where xxxxxx is the date in yyddmm format.
 2. Load 150 μ L into each standard well in duplicate.
 3. Load 150 μ L into each samples well in duplicate.
 4. Put in plate reader and press start.

Reading the Plate:

- 3
 - *Record absorbance at 532 nm.*
 1. Turn on Multiskan and open your saved file TBxxxxxx.sed.
 2. Place plate onto Multiskan holder and click **START**.
 3. Select Process>Curve Fit. Choose the appropriate data (usually Measure1), then click **OK**.
 4. Save Curve Fit data sheet as an Excel file into the Data folder/TBARS data folder. Use the naming convention TBxxxxxx.xls, where xxxxxx is the date in yymmdd format.