

Sep 20, 2018

## Degenerated PCR with GoTaq Hot Start

 [PLOS Genetics](#)

DOI

[dx.doi.org/10.17504/protocols.io.szyef7w](https://doi.org/10.17504/protocols.io.szyef7w)

Eduardo Dupim<sup>1</sup>, Antonio Bernardo de Carvalho<sup>1</sup>

<sup>1</sup>Universidade Federal do Rio de Janeiro

 Eduardo Dupim

Universidade Federal do Rio de Janeiro

OPEN  ACCESS



DOI: [dx.doi.org/10.17504/protocols.io.szyef7w](https://doi.org/10.17504/protocols.io.szyef7w)

External link: <https://doi.org/10.1371/journal.pgen.1007770>

**Protocol Citation:** Eduardo Dupim, Antonio Bernardo de Carvalho 2018. Degenerated PCR with GoTaq Hot Start. [protocols.io](#) <https://dx.doi.org/10.17504/protocols.io.szyef7w>

**Manuscript citation:**

Dupim EG, Goldstein G, Vanderlinde T, Vaz SC, Krsticevic F, Bastos A, Pinhão T, Torres M, David JR, Vilela CR, Carvalho AB (2018) An investigation of Y chromosome incorporations in 400 species of *Drosophila* and related genera. PLoS Genet 14(11): e1007770. doi: [10.1371/journal.pgen.1007770](https://doi.org/10.1371/journal.pgen.1007770)

**License:** This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working

**We use this protocol and it's working**

**Created:** August 27, 2018

**Last Modified:** September 20, 2018

**Protocol Integer ID:** 15128

**Keywords:** PCR, Degenerated PCR

## Abstract

Degenerated PCR to test Y-linkage of genes in several Drosophila species. The reactions are made separately for males and females of 400 species and subspecies of Drosophila and related genera.

## Materials

### MATERIALS

☒ GoTaq(R) Hot Start Polymerase, 500u **Promega Catalog #M5005**

☒ dNTP Mix, 10mM, 1000ul **Promega Catalog #U1515**

☒ DEPC-Treated Water **Ambion Catalog ##AM9906**

## Pre-Mix Preparation

- Usually, we performing PCR tests in large-scale, testing several DNA samples from different species at once. We prepared a pre-mix stock to economy time in PCR experiments.

Reagent	1 reaction	1000 reactions
D E P C - T reat e d W ater	1 1 . 6 uL	1 1 . 6 m L
5 x G re e n G o T aq Fl exi B uff er	4 0 uL	4. 0 m L
M g C l <sub>2</sub> 2.5 mM	2 0 uL	2. 0 m L

dNTP	0.4 μL	0.4 μL
10 mM M		
TOTAL VOLUME	18 μL	18 μL

We divide the pre-mix solution in 1 mL aliquots and stocked at -20°C.

## Final Degenerated PCR preparation

- 2 Normally, the DNA template concentration is 10 ug/uL or higher.

Reagent	1 reaction (20.1 uL)
Template	1 uL
Forward degenerated primer 40mM	0.5 uL
Reverse degenerated primer 40mM	0.5 uL
Premix	18 uL
GoTaq Hot Start Polymerase	0.1 uL

## PCR Programs

- 3 We used different thermocycle programs, according to the primers. In all programs, the GoTaq Hot Start Polymerase was previous incubated for 2 minutes to be activated. The PCRs were performed in a Applied Biosystems Veriti™ 96-Well Thermal Cycler (Cat#4375786).

**1) Degenerated PCR Program:** Differently of the normal PCR thermocycler programs, the degenerated PCR have more time for annealing.

cycles	Denaturat	Annealing	Polymerizat
1 x	95°C, 2:00 min	--	--
40 x	95°C, 30 min 0:30 min	x° C, 1:10 min / 0:00 pbof templat	72°C, 0:00 min
1 x	--	--	72°C, 7:00 min

x°C = optimal annealing temperature for the pair of primers.

**2) Degenerated Touchdown PCR (TD-PCR) Program:** In TD-PCR, we screen a range of annealing temperatures to try optimize the reaction in different species samples. So, we have a stage where the annealing temperature decrease -0.2°C by cycle, in the end of this stage, the annealing temperature decreased -4°C.

cycles	Denaturation	Annealing	Polymerization	
1 x	95°C, 2:00 min	---	---	
20 x	95°, 0:30 min	X° C, C, 1: 30 (Δ b y c y cl e )	72° C, 1:00 min / -0.2° 000 p b o f te m pl at e	
25 x	95°, 0:30 min	X° C, 0:30 min	72° C, 1:00 min / 1000 p b o f	

				te m pl at e
1 x	---	---		7 2 ° C, 7: 0 0 m in

x°C = optimal annealing temperature for the pair of primers.

### References:

- Sambrook, J. & Russell, D. W. (2001) 'Chapter 8. Protocol 11. Mixed Oligonucleotide-primed Amplification of cDNA' in *Molecular cloning : a laboratory manual*. Cold Spring Harbor: Cold Spring Harbor Laboratory, p. 8.66-8.71.