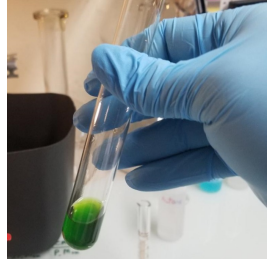


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Chlorophyll Extraction and Spectral Analysis V.1

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Protocol status: Working

We use this protocol and it's working

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Abstract

This protocol is designed to be able to extract and analyze the concentration of chlorophyll within a sample of a given plant. The procedures of this protocol require using a spectroscope to determine the approximate level a chlorophyll within a given sample.

Guidelines

For proper extraction and spectroscopy of Chlorophyll concentration you must be able to measure and transfer liquids within a hundred micro-liters ensure that samples are separated and free of contaminants.

Materials

MATERIALS

☒ Magnesium Sulfate Heptahydrate, ACS Grade **Gold Biotechnology Catalog #M-020**

☒ Acetone **Sigma Aldrich Catalog #34850**

STEP MATERIALS

☒ Magnesium sulfate, heptahydrate, ACS **Bio Basic Inc. Catalog #MB0329.SIZE.2.5Kg**

☒ Acetone **Sigma Aldrich Catalog #34850**

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Protocol materials

☒ Magnesium sulfate, heptahydrate, ACS **Bio Basic Inc. Catalog #MB0329.SIZE.2.5Kg**

In Materials, Materials and [2 steps](#)

☒ Magnesium Sulfate Heptahydrate, ACS Grade **Gold Biotechnology Catalog #M-020** Materials

☒ Acetone **Merck MilliporeSigma (Sigma-Aldrich) Catalog #34850**

In Materials, Materials, Materials, Materials and [3 steps](#)

Safety warnings

- ⚠ This protocol requires the use of flammable solvents, and require the extraction of pigments that may stain clothing. Proper lab coat, eye protection, gloves in ventilation are required to conduct this chlorophyll extraction and concentration protocol. Also, care must be taken to ensure that all materials used are disposed properly, as many of the chemicals may be hazardous to health and environment.

Before start

In order to perform this chlorophyll extraction protocol you will need the following materials and chemicals:

the materials listed are based on one single sample, in must be multiplied based on the number of samples you would like to test

One 20 ml (minimum) test tube

40 ml beaker

Two 200 micro-liters PCR tubes (although more may be needed based on the accuracy necessary for the procedures performed)

A 100 to 1000 micro-liter adjustable pipette

5 disposable 1000 micro-liter pipette tips (Number varies based on need and mistakes)

1200 g Centrifuge

One 20 micron filtration filter paper

Mortar and Pestle

Transfer Pipettes (as needed for contamination prevention)

50 ml Graduated Cylinder

Test tube stirrers

Approximate protocol time: 1 hours total

As needed Distilled water


15 ml Acetone



0.13g Magnesium sulfate



0.25g of sample


Spectrometer



Extraction of Chlorophyll



- 1 Weigh out  0.25 g of sample subject and add it to a pestle. 1m

- 2 Weigh out  0.13 g of
 Magnesium sulfate, heptahydrate, ACS **Bio Basic Inc. Catalog #MB0329.SIZE.2.5Kg** and add it to a pestle. 1m

- 3 Add  1.0 mL of
 Acetone **Sigma Aldrich Catalog #34850** 1m
to the pestle.

- 4 Grind the entire mixture until the sample is consistent and the
 Magnesium sulfate, heptahydrate, ACS **Bio Basic Inc. Catalog #MB0329.SIZE.2.5Kg** is completely dissolved into the sample paste. 5m

- 5 Add the paste into a 40 ml beaker and wash out the pestle with  1.0 mL of
 Acetone **Sigma Aldrich Catalog #34850** 1m
and empty it into the beaker.

- 6 Add  13.0 mL of
 Acetone **Sigma Aldrich Catalog #34850** 2m
to the 40 ml beaker and mix the sample thoroughly, ensuring to press and remix the sample several times.


- 7 Let the sample beaker stand for 15 minutes. 15m

- 8 Filter the sample into a 20 ml test tube using a 20 micron retention filter paper. 1m


- 9 Let the sample stand for a further 10 minutes. 10m



Sample Analyzation

10 Add  200 μL of the sample to two PCR tubes and label them according to your needed labeling system. the two tubes will serve as double tests for verification. 1m

11 Centrifuge the two samples to 1200 g to settle out any particulates for 1 minute. 1m

12 From each PCR tube, transfer  100 μL of the sample from the top to another PCR Tube. 2m

13 Place each tube in the spectrometer. 1m

14 Take to log base 10 of the absorbance percentage of the wavelength 647 nm and 664.5 nm and denote them as A_{647} and A_{664} . 5m

15 To calculate the concentration... 5m

$$\text{CHL}_A = 20.47A_{647} - 4.73A_{664}$$

$$\text{CHL}_B = 12.63A_{664} - 2.52A_{647}$$

$$\text{CHL}_{\text{Total}} = \text{CHL}_A + \text{CHL}_B$$

Divide each concentration by 1000, multiply by the acetone used, then divide by the sample mass to get the mg/g of sample

CITATION

Inskeep, William P., and Paul R. Bloom. (1985). Extinction Coefficients of Chlorophyll a and b in N,N-Dimethylformamide and 80% Acetone.. *Plant Physiology*, vol. 77, no. 2, Jan. 1985.

LINK

[doi:10.1104/pp.77.2.483](https://doi.org/10.1104/pp.77.2.483)

16 Confirm readings match to within an acceptable margin of error. 5m



Citations

Step 15

Inskeep, William P., and Paul R. Bloom.. Extinction Coefficients of Chlorophyll a and b in N,N-Dimethylformamide and 80% Acetone.

doi:10.1104/pp.77.2.483